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Gold-catalyzed glycosidations: synthesis of 1,6-anhydro saccharides

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1,6-Anhydro sugars are excellent synthons for the syntheses of medicinally important molecules^{[1](#page-2-0)} and materials.^{[2](#page-2-0)} They are also known to be precursors for the synthesis of proteoglycans, glycosyl halides, $3a$ N- or S-glycosides, $3b-d$ and C-glycosides. $3e$, Stereoregular or hyperbranched glycopolymers are also reported from 1,6-anhydro sugars by cationic ring-opening polymerization.[4](#page-2-0) Interesting opportunities for the synthesis of complex molecules and novel glycopolymers prompted the development of methods for the syn-thesis of this set of sugar derivatives.^{[5](#page-2-0)}

1,6-Anhydro sugar derivatives are synthesized either by thermal degradation or alternatively by chemical reactions.^{[5](#page-2-0)} Thermal degradation or pyrolysis exploited all the major conventional and nonconventional reaction media as well as the convection sources to get 1,6-anhydro sugars.^{5b–f} The major limitation of a thermal degradation process is that they are more suitable to 1,6-anhydro monosaccharides and really tough to limit pyrolysis for 1,6-anhydro oligosaccharides.5a However, 1,6-anhydro sugar derivatives by chemical reactions enjoy the control and selective synthesis which are otherwise not easy.^{1b} An usual sequence for the chemical synthesis of 1,6-anhydro sugars involves a leaving group at the anomeric position with the 6-hydroxyl group and the remaining hydroxyl groups are protected. Till date, Shoda's direct synthesis of 1,6-anhydro saccharides from unprotected glycopyranosides by the use of 2-chloro-1,3-dimethylimidazolinium chloride is the best.^{5a} In summary, most of the reported methods use stoichiometric quantities of reagents, often long reaction times, and tedious isolation procedures. Thus methods that enable the synthesis of 1,6-anhydro sugar derivatives through catalytic means are needed.^{1b}

The foregoing discussion encouraged us to ponder upon developing a catalytic route to the synthesis of 1,6-anhydro saccharides taking the cue from recent observations in gold-catalyzed glycosidations. In our laboratory, we have identified propargyl and methyl glycosides as novel glycosyl donors taking advantage of both the Lewis as well as the Brønsted acidity of $Au(III)$ salts.⁶ Usually, a glycosylation reaction involves a fully protected glycosyl donor with a leaving group at the anomeric position and a glycosyl acceptor (aglycone) that frequently contains a single hydroxyl moiety and the reaction happens in an intermolecular fashion.⁷

Thus we hypothesized that if the glycosylation is carried out intramolecularly on a sugar substrate containing a 6-OH group (other hydroxyl groups being protected) and a leaving group at the C-1 position, the reaction shall proceed to yield 1,6-anhydro sugars. This proposition was further fueled by a recent observation that showed cleavage of the interglycosidic bond and formation of 1,6-anhydro mannoside (4) and disaccharide 3 from an armed disaccharide (1) and aglycone 2 under gold-catalyzed glycosidation conditions ([Scheme 1](#page-1-0)). 8

Accordingly, an acetonitrile solution of propargyl 2,3,4-tri-Obenzyl mannopyranoside 5a was heated to 70 \degree C for 10 h to isolate 1,6-anhydro mannoside 4 in 71% yield after concentration in vacuo followed by filter column chromatographic purification. $9-11$ In another reaction, methyl mannopyranoside (5b) was also found to give compound 4 in 62% yield when subjected to above delineated reaction conditions [\(Scheme 2\)](#page-1-0).^{[10](#page-2-0)}

Aforementioned results enticed us to evaluate the general applicability of the current protocol for the synthesis of other 1,6-anhydro sugars of monosaccharides, disaccharides, and trisaccharides. 1,6-Anhydro sugar formation was then checked with $gluco-(6a,6b)$ and galacto- $(7a,7b)$ substrates as well to obtain the corresponding 1,6-anhydro derivatives 8 and 9 in good yields ([Table 1](#page-1-0)).⁹ In addition, the suitability of 1,6-anhydro sugar formation has been studied with a panel of disaccharides and trisaccharides. Interesting to note that AuBr₃-catalyzed intramolecular reaction occurred on disaccharides (10a, 10b, 11a, and 11b) and trisaccharides (14a, 14b, 15a, and 15b) resulting in the formation of the corresponding 1,6-anhydro sugar derivatives (12, 13, 16, and 17) in good yields.^{[9](#page-2-0)} It is pertinent to mention that the inherent acidity in the Au(III)-catalyzed glycosidation showed the domino effect in one-pot. Deprotection of 6-O-silyl ether happened first to give the 6-OH compound that got subsequently converted to

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Scheme 1. Cleavage of interglycosidic bond and formation of 1,6-anhydro mannoside.

1,6-anhydro sugar via the intramolecular trapping of the oxocarbenium ion generated due to the extrusion of the anomeric alkyl (propargyl/methyl) group by the action of AuBr₃.

In conclusion, the synthesis of 1,6-anhydro sugars from the corresponding 6-hydroxy propargyl/methyl glycosides was realized in the presence of a catalytic amount of $AuBr₃$ at 70 °C in acetonitrile. Interestingly, deprotection of 6-O-TBDPS ether was observed for the first time in the presence of catalytic quantity of $AuBr₃$ and subsequent intramolecular glycosidation happened in domino

Scheme 2. AuBr₃ catalyzed synthesis of 1,6-anhydro mannoside from propargyl and methyl mannopyranosides.

Table 1 Synthesis of 1,6-anhydro saccharides

Substrate	Product	Time, Yield	
		$R = \lambda_2$	$R = CH3$
OH BnO ² BnO BnO R 6a R = $\sqrt[3]{x^2}$ \gg , 6b R = CH ₃	O OBn O. OBn OBn 8	4 h, 75%	24 h, 69%
BnO BnO R \mathbb{R} , 7b R = CH ₃ $7a R =$ Wy	OBn BnO OBn 9	12 h, 76%	18 h, 65%
OTBDPS BnO BzO BzO ² BzO Ŕ. OBz	OBn OBn BzO ⁻ BZO BZO OBz	18 h, 71%	20 h, 64%
10a R = $\sqrt[n]{x_i}$, 10b R = CH ₃ OBz OTBDPS BzO ² BnO BzO BnO^{\dagger}_{O} R \gg , 11b R = CH ₃ 11a R = $v_{\rm c}$	12 OBn OBz BzO ² BzO OBn OBz 13	20 h, 72%	24 h, 68%
OTBDPS OBz BnO Rn(BzO BzO R Bz _O OBz OBz	OBn OBz BzO OBn BZO BzO $\frac{1}{\text{O}Bz}$ OBz	20 h, 67%	18 h, 64%
14a R = $\sqrt[n]{x_i}$, 14b R = CH ₃ OBz OBz OTBDPS BzQ BzO \overrightarrow{OBz} OBZ $BnO_0 \rightarrow R$ \gg , 15b R = CH ₃ 15a R = $\sqrt[n]{2}$	16 OBn OBz OBz BzO O OBn BzO OBZ OBZ OBz 17	10 h, 65%	18 h, 62%

fashion to give 1,6-anhydro sugar derivatives of disaccharides and trisaccharides.

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Supplementary data

Supplementary data associated with this article can be found, in the online version, at [doi:10.1016/j.tetlet.2010.09.004](http://dx.doi.org/10.1016/j.tetlet.2010.09.004).

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9. All products gave satisfactory ¹H, ¹³C, DEPT NMR and MS analysis. See Supplementary data.
- 10. General procedure for the synthesis of 1,6-anhydro sugars: To a solution of 6 hydroxy alkyl glycopyranoside (0.1 mmol) in anhydrous acetonitrile (2 mL) was added 7 mol % of AuBr₃ under argon atmosphere at room temperature. The resulting mixture was heated to 70° C and stirred till the completion of the reaction as judged by TLC analysis ([Table 1\)](#page-1-0). The reaction mixture was concentrated in vacuo to obtain a crude residue which was purified by silica gel column chromatography using ethyl acetate–petroleum ether (1:4) as mobile phase.
- 11. (a) Compound characterization data for compound 4: α_{I}^{25} (CHCl₃, c 1.0) –16.6;
¹H NMR (200.13 MHz, CDCL); λ 3.47(t, 1H J 1.8 Hz), 3.58(dd, 1H J 1.8 ¹H NMR (200.13 MHz, CDCl₃): δ 3.47(t, 1H, J = 1.8 Hz), 3.58(dd, 1H, J = 1.8, 5.4 Hz), 3.73(dd, 1H, J = 6.0, 7.1 Hz), 3.81(qd, 1H, J = 1.6, 3.1, 5.0 Hz), 4.25(dd, 1H, J = 0.9, 7.1 Hz), 4.43–4.57(m, 5H), 4.52(ABq, 2H, J = 12.4 Hz), 5.46(s, 1H),
7.20–7.38(m, 15H); ¹³C NMR (50.32 MHz, CDCl₃): δ 65.0, 71.3, 71.4, 73.4, 74.1, 74.4, 74.5, 76.5, 100.1, 127.7–128.5, 137.6, 137.9, 137.9; HRMS (MALDI-TOF) Calcd for $C_{27}H_{28}O_5$ Na: 455.1834; Found, 455.1830.

(b) Compound characterization data for compound **8**: $[x]_2^{25}$ (CHCl₃, *c* 1.0) –26.0;
¹H NMB (200.13 MHz, CDCL): $\frac{3}{2}$ 3 3/(d, 2H, I – 1.7 Hz), 3.58(quintet, 1H, I – 2.4 H^1 H NMR (200.13 MHz, CDCl₃): δ 3.34(d, 2H, J = 1.7 Hz), 3.58(quintet, 1H, J = 2.4, 3.6 Hz), 3.67(dd, 1H, $J = 5.9$, 7.2 Hz), 5.91(dd, 1H, $J = 0.8$, 7.2 Hz), 4.41(ABq, 2H, J = 12.5 Hz), 4.48–4.63(m, 5H), 5.46(s, 1H), 7.21–7.35(m, 15H); ¹³C NMR $(50.32 \text{ MHz}, \text{ CDCl}_3): \delta 65.3, 71.1, 71.7, 71.9, 74.2, 75.8, 75.9, 76.6, 100.5,$ 127.5–128.4, 137.7, 137.8, 137.8; HRMS (MALDI-TOF) Calcd for C₂₇H₂₈O₅Na, 455.1834; Found, 455.1840.

(c) Compound characterization data for compound 9: $[x]_D^{15}$ (CHCl₃, *c* 1.2) –34.9; ¹H NMR (200.13 MHz, CDCl₃): δ 3.52(t, 1H, J = 1.7 Hz), 3.62(t, 1H, J = 6.0 Hz) 3.77–3.92(m, 2H), 4.35–4.66(m, 8H), 5.36(t, 1H, J = 1.7 Hz), 7.22–7.37(m, 15H);
¹³C NMR (50.32 MHz, CDCl₃): δ 64.3, 71.1, 72.1, 72.7, 73.0, 73.1, 74.1, 76.3
100.2, 127.6–128.5, 137.5, 137.9, 138.2; HRMS (MALDI-TOF) C27H28O5Na, 455.1834; Found, 455.1836.

(d) Compound characterization data for compound 12: $[\alpha]_D^{25}$ (CHCl₃, c 1.1) +16.2; ¹H NMR (200.13 MHz, CDCl₃): δ 3.07(s, 1H), 3.42(s, 1H), 3.49(dd, 1H, J = 5.9, 7.3 Hz), 3.77(dd, 1H, J = 0.8, 7.3 Hz), 3.88(m, 1H), 4.31-4.57(m, 7H), $4.62(ABq, 2H, J = 12.4 Hz)$, $5.21(s, 1H)$, $5.38(dd, 1H, J = 8.0, 9.6 Hz)$, $5.58(t, 1H, J)$ J = 9.8 Hz), 5.75(t, 1H, J = 9.8 Hz), 7.20–7.76(m, 22H), 7.77–8.09(m, 8H); ¹³C NMR (100.61 MHz, CDCl₃): δ 62.8, 64.8, 69.5, 71.2, 71.4, 72.2, 72.2, 72.5, 74.2, 75.2, 75.7, 76.0, 99.8, 100.2, 127.6–129.9, 133.2, 133.3, 133.3, 133.5, 137.7, 137.9, 164.9, 165.2, 165.7, 166.0; HRMS (MALDI-TOF) Calcd for C₅₄H₄₈O₁₄Na, 943.2942; Found, 943.2949.

(e) Compound characterization data for compound 13: $[\alpha]_D^{25}$ (CHCl₃, c 0.9) -13.4 ; ¹H NMR (500.13 MHz, CDCl₃): δ 3.27(s, 1H), 3.55(dd, 1H, J = 6.2, 7.2 Hz), $3.65(s, 1H)$, $3.80(s, 1H)$, $3.84(d, 1H, J = 7.3 Hz)$, $4.10(m, 1H)$, $4.41(ABq, 2H, 1H)$ $J = 12.0$ Hz), 4.42 (ABq, 2H, $J = 12.3$ Hz), 4.43 (dd, 1H, $J = 5.3$, 12.2 Hz), 4.56 (dd, $1H, I = 3.0, 12.1 \text{ Hz}$), $4.63(d, 1H, I = 5.8 \text{ Hz})$, $5.25(d, 1H, I = 5.8.1 \text{ Hz})$, $5.37(s, 1H)$, 5.56(dd, 1H, J = 8.1, 9.8 Hz), 5.66(t, 1H, J = 9.8 Hz), 5.99(t, 1H, J = 9.6 Hz), 7.15-
7.56(m, 22H), 7.80–8.05(m, 8H); ¹³C NMR (125.76 MHz, CDCl₃): δ 63.0, 65.4, 69.6, 71.2, 72.0, 72.1, 72.4, 73.0, 74.2, 75.3, 77.0, 77.7, 99.7, 100.5, 127.5–129.9, 133.2, 133.2, 133.3, 133.5, 137.6, 137.8, 165.0, 165.2, 165.8, 166.1; HRMS(MALDI-TOF) Calcd for C₅₄H₄₈O₁₄Na, 943.2942; Found, 943.2950.

(f) Compound characterization data for compound **16:** $[\alpha]_{D}^{25}$ (CHCl₃, *c* 1.3) +47.4; ¹H NMR (400.13 MHz, CDCl₃): δ 3.05(s, 1H), 3.33(m, 1H), 3.38(s, 1H), 3.49(t, 1H, *J* = 6.3 Hz), 3.65–3.98(m, 5H), 4.14(t, J = 8.1 Hz), 4.44(ABq, 2H, J = 12.2 Hz), 4.38–4.64(m, 5H), 4.71(m, 1H), 4.86(d,
1H, J = 7.8 Hz), 5.22(s, 1H), 5.31(dd, 1H, J = 8.1, 9.9 Hz), 5.44(dd, 1H, J = 3.3,
10.3 Hz), 5.64(t, 1H, J = 9.4 Hz), 5.78(m, 1H), 7.05–8.15(m $(125.76 \text{ MHz}, \text{CDCl}_3): \delta 61.0, 61.9, 64.6, 67.4, 69.9, 70.9, 71.2, 71.3, 71.6, 72.3,$ 72.5, 72.8, 73.8, 74.9, 75.0, 75.7, 76.2, 99.7, 100.2, 101.0, 127.6–130.0, 133.1, 133.2, 133.3, 133.4, 133.5, 133.6, 133.6, 137.6, 137.9, 164.7, 165.0, 165.2, 165.3, 165.4, 165.6, 165.7; HRMS(MALDI-TOF) Calcd for $C_{81}H_{70}O_{22}Na$, 1417.4256; Found, 1417.4251.

(g) Compound characterization data for compound 17: $[\alpha]_D^{25}$ (CHCl₃, c 1.2) +28.2; ¹H NMR (500.13 MHz, CDCl₃): δ 3.23(s, 1H), 3.54(dd, 1H, J = 6.1, 7.1 Hz), 3.58(td, 1H, J = 3.3, 6.1, 10.3 Hz), 3.63(dd, 1H, J = 6.7, 11.4 Hz), 3.66(m, 1H),
3.72(dd, 1H, J = 6.7, 11.3 Hz), 3.76–3.94(m, 2H), 4.18–4.63(m, 9H), 4.89(ABq, 2H, J = 7.7 Hz), 5.34(d, 1H, J = 1.1 Hz), 5.35(dd, 1H, J = 3.5, 10.6 Hz), 5.49(dd, 1H,
J = 8.1, 10.2 Hz), 5.69–5.77(m, 3H), 7.05–8.08(m, 45H); ¹³C NMR (125.76 MHz CDCl₃): δ 61.1, 62.2, 65.3, 67.5, 69.9, 71.0, 71.4, 71.8, 71.8, 71.9, 73.0, 73.1, 74.3, 75.0, 75.8, 76.0, 78.7, 100.4, 100.5, 101.0, 127.4–130.0, 133.1, 133.2, 133.3, 133.4, 133.4, 133.4, 133.5, 137.6, 139.9, 164.7, 165.0, 165.2, 165.4, 165.4, 165.6, 165.8; HRMS(MALDI-TOF) Calcd for $C_{81}H_{70}O_{22}Na$, 1417.4256; Found, 1417.4251.